

RESPONSE OF FUSARIUM MONILIFORME - THE CAUSAL ORGANISM OF BAKANAE DISEASE OF RICE AGAINST DIFFERENT FUNGICIDES

SACHIN KUMAR JAIN*, KAMAL KHILARI, MOHD ALI AND RANVIJAY SINGH

Department of Plant Pathology,

S. V. P. Uni. of Ag. and Tech Modipuram, Meerut - 250 110, INDIA e-mail: sachinjain1115@gmail.com

KEYWORDS

Rice, Bakanae Foot rot *Fusarium moniliforme* and fungicides

Received on : 21.01.2014

Accepted on : 25.02.2014

*Corresponding author

INTRODUCTION

ABSTRACT

Bakanae disease is becoming a serious threat to basmati production and had been reported to cause 20-50% loss in total production. Bakanae disease is caused by *Fusarium moniliforme* Sheldon. In the present investigation seven fungicides were evaluated *in vitro* condition for management of *Fusarium moniliforme*. Result was indicated that, carbendazim cent percent inhibited to pathogen growth at all concentration (1000ppm, 500ppm, 100ppm and 10ppm). All other systemic fungicides such as hexaconazole, tabuconazole and thiophanate completely inhibited the pathogen at 100ppm, 500ppm and 1000ppm but at 10ppm they inhibited pathogen growth 75.28%, 97.78% and 57.64% respectively. All other non systemic fungicides likes mancozeb, propineb and thiram were found less effective and at 1000ppm these inhibited pathogen growth as 45.67%, 57.40% and 46.53% respectively. Thus, the present study demonstrated the use of systemic fungicides have the good inhibition potential against *Fusarium moniliforme*.

Rice is an important cereal crop and growing all over the world. Rice grows in India is primarily divided into Basmati rice and Non-Basmati rice. India is the major producer and exporter of basmati rice to the world. Rice production is very much effected by many biotic and abiotic stresses. Among the biotic stress, fungal diseases have important role in reduction the yield of rice. Bakanae (Foot rot) is one of them that is emerging problem as major disease of rice. Bakanae disease is caused by Fusarium moniliforme Sheldon and the pathogen was later identified as F. fujikuroi Nirenberg. The teleomorph stage of F. moniliforme is known as Gibberella fujikuroi Sawada (Nirenberg, 1976). The teleomorph, Gibberella fujikuroi, has been reported on rice in China, Japan and Taiwan (Sun, 1975). The fungus produces gibberellins and other secondary metabolites such as carotenoids, bikaverin and fusarin, which directly affect the growth of rice, plants (Ilija et al., 2009). The fungus produces giberrellin hormone which causes elongation of plant (Nyvall, 1999). The pathogen is widely distributed through the world and has wide host range (Kazempour, 2007). Precise information on losses caused by Bakanae disease 15% was reported in India and 40-50% in Japan (Pavgi and Singh, 1964). The most visible symptoms on rice plant of F. fujikuroi are seedling elongation, foot rot, seedling rot, grain sterility, and grain discoloration (Ou, 1985). Seed treatment with fungicide such as thiram has been used for management of Bakanae disease (Suzuki et al., 1985). After the emergence of pathogen resistance to these fungicides, ipoconazole has become the major fungicide to control the disease (Kumazawa *et al.* 2000). In present investigation, commonly available fungicides were tested *in vitro* to known their toxicity on the pathogen because these may be important component in integrated disease management programme.

MATERIALS AND METHODS

Present investigation on *in vitro* evaluation of different fungicides against *Fusarium moniliforme* causing Bakanae disease of rice was carried out in the department of plant pathology, Sardar Vallabhbhai Patel University of Agriculture and Technology, Meerut and Uttar Pradesh.

Seven fungicides were tested *in vitro* against *F. moniliforme* by food poison technique (Nene and Thapliyal, 1971). Required amount of fungicide was added in sterilized potato dextrose media to obtain 10ppm, 100ppm, 500ppm, and 1000ppm concentration in the conical flask. Fungicide was mixed in potato dextrose media by shaking the flask prior to pouring in sterilized Petri plates. The medium was allowed to solidify and then 3 mm discs from seven days old culture of *F. moniliforme* was placed in centre of each Petri plate. Control was maintained without any treatment. Three replications were maintained for each concentration for every tested fungicide. Inoculated plates were incubated at $28 \pm 1^{\circ}$ C in BOD incubator. Observations were recorded on radial growth of test pathogen at regular interval. Percent inhibition over control was calculated by the following formula given below (Vincent,

1947).

$$I = \frac{C - T}{C} \times 100$$

where,

I = Percent inhibition of fungal growth

C = Radial growth in control

T = Radial growth of treated petridish

RESULTS AND DISCUSSION

The seven fungicides namely tabucanazole, thiophanate, carbendazim, hexaconazole, mancozeb, thiram and propineb were tested *in vitro* against the pathogen by the food poison technique at the 10 ppm, 100 ppm, 500 ppm, and 1000 ppm concentration. Table 2 and Fig. 1 indicates the results of different fungicides on pathogen at different concentrations.

It was recorded that at 10 ppm concentration, carbenbazim completely inhibited the growth of pathogen followed by tabuconazole (97.78%), hexacanazole (75.28%) and thiophanate (57.64%) while Mancozeb, thiram and propineb expressed minimum growth inhibition that is 0.97, 1.84 and

Table 1: Fungicides tested against the F. moniliforme

Fungicides	Chemical composition
Tebuconazole	(RS)-1-p-chlorophenyl-
	4, 4-dimethyl-3-(1H-1,2,4-triazol-1-ylmethyl)
	Pentan-3-ol
Carbendazim	Methyl benzimidazol-2-ylcarbamate
Thiophanate	Diethyl4,4-(o-henylene)bis (3- thioallphanate
Hexaconazole	2-(2, 4-dichlorophenyl)-1-(1H-1, 2, 4-triazol-
	1-yl) hexan-2-ol
Mancozeb	Manganese ethyl
	bisdithiocarbamate
Propineb	zinc propylenebisdi thiocarbamate
Thiram	Tetramethyl thiuram Disulfide

0.73 respectevly. Tabuconazole, thiophanate, and hexacanazole shown complete inhibition of pathogen growth at 100ppm while mancozeb, propineb and thiram shown inhibition as 5.91, 10.26 and 4.93 respectively. At the 500ppm mancozeb inhibited the growth of pathogen 41.86% followed by propineb, 20.51% and thiram 4.71%. Growth inhibition at 1000ppm was observed 45.67, 57.40 and 46.53 in case of mancozeb, propineb and thiram respectively. Previous studies also found that systemic fungicide are best for management of Bakanae disease in laboratory as well as in field. Biswas and Das (2002) tested the fungicides at 0.2% and found that, carbendazim and benomyl significant reduce Bakanae disease incidence and percentage tiller infection, and increase in yield. Titone et al. (2004) found that, fungicide like carbendazim, mancozeb, iprodione + triticonazole and carboxin + thiram reduced the Bakanae disease incidence by 80 and 40% under laboratory and field conditions respectively. Taskeen-un-Nisa et al. (2011) tested the efficacy of carbendazim, hexaconazol, bitertanol, myclo-butanil, mancozeb, captan and zineb against *Fusarium* oxysporum and found that, the maximum inhibition in mycelial growth was observed in the hexaconozole at 1000 ppm followed by other fungicides at the same concentration. Kapadiya (2013) tested the systemic and non systemic fungicides against the Fusarium solani in vitro condition and found that among the systemic fungicide tebuconazole and carbendazim gave cent percent growth inhibition.

In the conclusion we suggested that use of chemical compounds may be the important component for management of Bakanae disease of rice. The present finding demonstrated the all systemic fungicides (tabucanazole, thiophanate, carbandazim, hexacanazole) as effective component in integrated disease management.

ACKNOWLEDGMENT

We would like to extend our sincere thanks to HOD of department of Plant Pathology, S. V. P. University of Agriculture

Table 2: Colony radial growth and percent inhibition of *F. moniliforme* on different fungicides

Treatment	10ppm *Average Colony diameter (mm.)	Percent inhibition (%)	100ppm *Average Colony diameter (mm.)	Percent inhibition (%)	500ppm *Average Colony diameter (mm.)	Percent inhibition (%)	* Average Colony diameter (mm.)	1000ppm Percent inhibition (%)
Tabucanazole 5% SC (Raxil)	2.00	97.78	0.00	100	0.00	100	0.00	100
Carbendazim 50% WP (Agrizim)	0.00	100	0.00	100	0.00	100	0.00	100
Thiophanate Methyl 70% WP (Prism)	38.12	57.64	0.00	100	0.00	100	0.00	100
Hexacanazole 5E (Cartaf)	22.24	75.28	0.00	100	0.00	100	0.00	100
Mancozeb 75% WP (Indosal M-45)	89.12	0.97	84.68	5.91	52.32	41.86	48.89	45.67
Propineb 70% WP (Antracol)	89.34	0.73	80.76	10.26	71.54	20.51	38.34	57.4
Thiram (Thiram)	88.34	1.84	85.56	4.93	85.76	4.71	48.12	46.53
Control C.D. at 5%	90 0.556	0.00	90 0.462	0.00	90 0.341	0.00	90 0.35	0.00

*Means of three replication

RESPONSE OF FUSARIUM MONILIFORME



500ppm



 T_1 - Tabucanazole, T_2 - Carbendazim, T_3 - Thiophanate, T_4 - Hexacanazole, T_5 - Mancozeb, T_6 - Propineb, T_7 - Thiram, T_8 - Control Figure 1: evaluation of fungicides against growth of *F. moniliforme (In vitro)*

and Technology, Meerut for providing the research facilities.

REFERENCES

Biswas, S. and Das, S. N. 2002. Efficacy of fungicides for the control of Bakanae disease of rice. Ann. Pl. Prote. Sci. 10(2): 288-290.

Ilija, K. K., Sasa, K. M. and Emilija, D. K. 2009. *Gibberella fujikuroi* (Sawada) Wollenweber, the new parasitical fungus on rice in the Republic of Macedonia. pp.175-182.

Kapadiya, I. B., Akbari, L. F., Siddhapara, M. R. and Undhad, S. V. 2013. Evaluation of fungicides and herbicides against the root rot of okra. *The Bioscan.* 8(2): 433-436.

Kazempour, M. N. and Elahinia, S. A. 2007. Biological control of *Fusarium fujikuroi*, the causal agent of Bakanae disease by rice associated antagonistic bacteria. *Bulgarian J. Agric. Sci.* **13(4):** 393-408.

Kumazawa, S., Ito, A., Saishoji, T. and Chuman, H. 2000. Development of new fungicides, ipconazole and metconazole. *J. Pestic.* Sci. 25: 321-331.

Nene, Y. L. and Thapliyal, P. N. 1971. Fungicides in Plant diseases control. Oxford and IBH publications Co. Pvt. Ltd. New Delhi. pp. 537-540.

Nirenberg, H. I. 1976. Untersuchungen uber die Morphologische and Biologische Differenzierung in Fusarium - Sektion Liseola. *Mitt. Biol. Bundesansi. Land-Forstwirtsch. Berlin - Dahlem.* 169: 1-117.

Nyvall, R. F. 1999. Field Crop Diseases. *Iowa State University Press, USA*. p. 1,021

Ou, S. H. 1985. *Rice Diseases,* p: 380. Great Britain (UK): Commonwealth Mycological Institute International Rice Research Institute. *Field Problems of Tropical Rice.* p. 172.

Pavgi, M. S. and Singh, J. 1964. Bakanae and foot rot of rice in Uttar Pradesh, India. *Plant Disease Reporter*. 48: 340-342.

Sun, S. K. 1975. The diseases cycle of rice Bakanae disease in Taiwan, Proc. Natl. Sci. Counc. Repub. 8: 245-255.

Suzuki, H., Takahashi, S., Fujita, Y. and Sonoda, R. 1985. Effect of seed disinfection by thiram benomyl on blast, brown spot and "Bakanae" disease of rice (in Japanese). *Ann. Rep. Plant Prot. North Jpn.* **36**: 122-124.

Taskeen, A. H., Bhat, M. Y., Pala, S. A. and Mir, R. A. 2011. In vitro inhibitory effect of fungicides and botanicals on mycelial growth and

spore germination of Fusarium oxysporum. J. Biopesticides. 4: 1.

Titone, P., Grassi, G., Polenghi, G., Tamborini, L. and Garibaldi, A. 2004. Rice seed dressing:chemical and thermal treatments to control Bakanae disease [Italian]. *Informatore Fitopatologico*. 54(5): 41-49.

Vincent, J. M. 1947. Distortion of fungal hyphal in the presence of certain inhibitors. *Nature*. 159: 850-853.